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1. Data representation and transformation

- Let I_t (treated cell) and I_r (reference cell) be the fluorescence signal intensities measured from each spot "i" on a microarray.
- Transform the data from a "cartesian" representation to a "polar" form, i.e.:

$$\begin{aligned} \text{Cartesian form } (I_t, I_r) &\rightarrow \text{Polar form } (R, x_i) \\ R_i &= \sqrt{I_t^2 + I_r^2} \quad (\text{intensity magnitude}) \\ x_i &= \frac{I_t}{I_t + I_r} \quad (\text{fractional intensity}) \end{aligned}$$

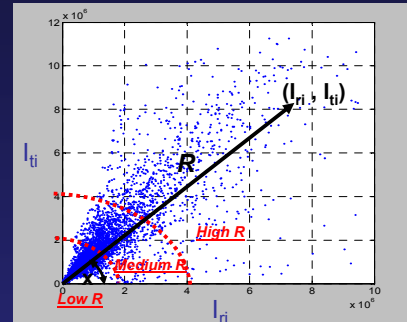


Figure 1. Intensity signal of control versus treated cell.

The data can be partitioned into at least 3 groups:
- Low R (see Fig. 1)
- Medium R
- High R

If intensity signal \propto (# of mRNA molecules)
Intensity measurements (I) \sim Gamma distribution
 $x_i = \frac{I_t}{I_t + I_r} \sim$ Beta distribution

Advantages of the polar transformation:

It is more natural (the variance of the microarray data changes with intensity measurement).

It normalizes the data more efficiently because none of the original information is lost

It enables statistical rigor and endows the problem with analytical tractability not possible with either raw intensity data or the popular intensity ratios.

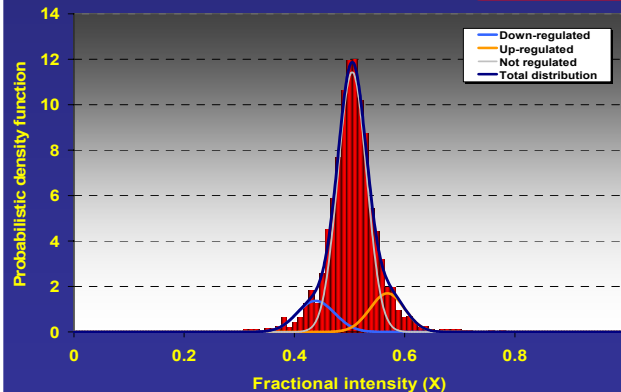


Figure 2. Histogram of fractional intensities and fitting of Beta distributions.

2. Drawing inferences

(i) Bias detection and correction

Theoretically, if there is NO bias the mean of $f_0(x)$ should be $\mu_0 = 0.5$

Bias is indicated if $\mu_0 \neq 0.5$; remove via correction for x_i :

$$\tilde{x}_i = \frac{x_i}{x_i + w(1-x_i)}$$

where $w = \mu_0 / (1 - \mu_0)$

The histogram of fractional intensity (X) data can be characterized as a mixture of at least three Beta distribution functions:

$$f(x) = \phi_L \cdot f_L(x) + \phi_0 \cdot f_0(x) + \phi_H \cdot f_H(x)$$

$\sum \phi_i = 1$

Where,

$f_L(x)$ = Beta distribution function for genes showing lower differential expression.

$f_0(x)$ = Beta distribution function for genes showing no differential expression.

$f_H(x)$ = Beta distribution function for genes showing higher differential expression.

(ii) Probability of expression status

Using the probability distribution functions it is possible to calculate the probability of expression status (down, up or not regulated) for every gene.

Example: The probability that gene g_i with a fractional intensity x_i is down-regulated (D) is given by:
 $P(g_i \in D) = \frac{\phi_D f_D(x_i)}{\phi_D f_D(x_i) + \phi_0 f_0(x_i) + \phi_U f_U(x_i)}$

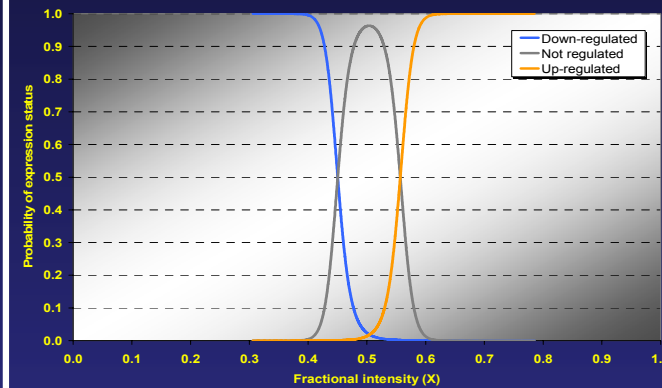


Figure 3. Probability of expression status.

Table 1 shows the identification of relevant genes using classical fold-change criteria. These results are compared before and after bias correction using our probabilistic framework.

Table 2 shows the number of candidates genes identified per group (low, medium and high R). All of them with a very high probability of expression status.

Figure 2 shows the probability of expression status (down, up or not differentially regulated) for each gene as a function of the fractional intensity (X).

3. Application results

Experimental data from gene expression studies in *Deinococcus radiodurans* following DNA damage (carried out by Dr. John Battista of LSU).

Table 1. "Fold-change" criterion results.

	2 Fold-change	3 Fold-change
Total genes identified	743	192
Up-regulated	145	77
Down-regulated	598	115
With Bias correction		
Total genes identified	429	131
Up-regulated	233	104
Down-regulated	196	27
False positives	402	88
False negatives	88	27

Table 2. Probabilistic framework results.

	Count	x value Thresholds	Equivalent "Fold change"	Associated Probabilities
(High R group)				
Total genes identified	68			
Up-regulated	68	$x \geq 0.83$	4.74	prob \approx 1.0
Down-regulated	N/A	N/A	N/A	N/A
(Med R group)				
Total genes identified	7			
Up-regulated	2	$x \geq 0.81$	4.18	prob = 1
Down-regulated	5	$x \leq 0.28$	0.38 (2.64)	prob = 1
(Low R group)				
Total genes identified	17			
Up-regulated	6	$x \geq 0.74$	2.85	prob = 1
Down-regulated	11	$x \leq 0.28$	0.38 (2.62)	prob \approx 1

4. Future work

Extend to GeneChip® technology.

Compare different microarray technologies; develop appropriate metrics for assessing the "quality".

Incorporate degree of confidence in the inference regarding the probability of expression status.